Engineering restoration and gaseous carbon uptake on a degraded bog: the role of *Eriophorum angustifolium* and micropropagated *Sphagnum*

Anna T. Keightley¹, Chris D. Field¹, James G. Rowson², Neal A. Wright³, Simon J.M. Caporn¹

¹ Department of Natural Sciences, Manchester Metropolitan University, Manchester, UK ² Jacobs UK Limited, Manchester, UK ³ Micropropagation Services (EM) Ltd (BeadaMoss®), East Leake, Loughborough, UK

SUMMARY

Degraded peatlands are significant sources of carbon greenhouse gases (CGHG), and their recovery can make significant contributions to climate change mitigation as well as deliver biodiversity benefits. Sphagnum mosses are key species for northern peatland formation and re-introduction is often needed for successful ecohydrological restoration of degraded bogs, but natural sources are scarce and often protected. Micropropagated Sphagnum moss products (BeadaMoss®) were developed to alleviate this constraint. This research explored in detail, for the first time, the CGHG fluxes on a cut-over lowland peatland restoration site where micropropagated Sphagnum was introduced to an existing 'nurse crop' of Eriophorum angustifolium, and tested the influence of vegetation maturity. Ecosystem CGHG flux was measured using closed chambers at plot scale in areas of both mature and immature E. angustifolium with and without application of BeadaGelTM Sphagnum, with control plots on bare peat. Studies were conducted over two years of contrasting weather patterns. In Year 1, mean net (CO₂e) CGHG uptake on vegetated plots was -2.33 (minimum 1.55, maximum -5.55) t ha⁻¹ yr⁻¹ with increasing CGHG uptake as vegetation matured. In Year 2, gross photosynthesis reduced significantly during the 2018 summer drought resulting in a small mean net CGHG emission of 0.11 (minimum 2.21 maximum -1.22) t ha⁻¹ yr⁻¹. Sphagnum application within immature vegetation resulted in greater CGHG uptake in both years, but was not as beneficial within mature vegetation. CGHG emission from bare peat (3.79 t ha⁻¹ yr⁻¹ overall) showed the magnitude of avoided losses. Methane flux contributed significantly to CGHG emission but was not closely related to water table depth. Application of Sphagnum within E. angustifolium can deliver good CGHG flux results in the early stages of degraded lowland bog recovery but cannot fully mitigate vulnerability to climate change scenarios.

KEY WORDS: climate change, cottongrass, greenhouse gases, methane, peatlands

INTRODUCTION

Although peatlands cover only 3 % of Northern Hemisphere land (Gorham 1991) they are estimated to contain 500 ± 100 Gt of carbon (C) or around one third of the world's soil carbon (Yu 2012), which is more C than in all forests globally (Joosten et al. 2016). However, an estimated 15 % of the world's peatlands are drained and damaged (Joosten et al. 2012) making them a source of carbon dioxide (CO_2) emissions (Leifeld & Manichetti 2018) that, with peat fires, accounts for around 5 % of all anthropogenic CO₂ emissions (Crump 2017). This amounts to ~2 Gt yr⁻¹ (Joosten 2016), which was double the global CO₂ emissions from aviation at that time (Graver et al. 2019). Methane (CH₄) emission from peatlands is also seen as a significant contributor to climate forcing (Glatzel et al. 2004, van Winden et al. 2012, Haddaway et al. 2014, Turetsky et al. 2014), and Evans et al. (2017) currently estimate that natural UK peatlands are 'climate neutral' owing to climate forcing of CH₄ emissions counteracting CO₂ uptake.

Peatland restoration can modify emissions of carbon greenhouse gases (CGHG; i.e. CO₂ and CH₄) (Waddington & Warner 2001, Aitova et al. 2023) and is seen as an achievable climate change mitigation activity (Bain et al. 2011, Joosten et al. 2012), but rapid implementation is essential to achieve the greatest benefit (Glenk et al. 2021). Rewetting is a key factor in encouraging the recovery of vegetation and protecting carbon stocks in degraded peatlands (Joosten et al. 2012, González & Rochefort 2014). Günther et al. (2020) explore the dichotomy between CO₂ emission from drained peatlands and the CH₄ emission from rewetted peatlands and conclude that "CH₄ radiative forcing does not undermine the climate change mitigation potential of peatland rewetting". Sphagnum mosses may reduce CH4 flux due to the presence of methanotrophic bacteria which oxidise methane for use by the plant, although this



may be temperature or water table depth (WTD) dependent (Kip *et al.* 2010, Larmola *et al.* 2010, van Winden *et al.* 2012). The optimum water table level to limit both CO_2 and CH_4 losses is reported to be 10 cm below the surface (Evans *et al.* 2021).

In the UK, the techniques and benefits of lowland peatland restoration are presently under-researched compared to upland systems (Haddaway et al. 2014) and there is currently a lack of data on CGHG fluxes from degraded lowland raised bogs (Evans et al. 2017). Sphagnum mosses are an intrinsic part of lowland bog formation and development in the hemisphere, bioengineer the northern and continuously environment for favourable ecohydrological conditions (van Breemen 1995, Quinty & Rochefort 2003) through chemical processes (Rydin & Jeglum 2013) and buffering against evapotranspiration (Mazziotta et al. 2019). The primary aim of lowland peatland restoration is to develop a Sphagnum-dominated acrotelm of bog vegetation that stabilises the water table, protecting current peat stocks in the short term and promoting peat and carbon accumulation in the long term (Lindsay 2010, Lucchese et al. 2010, Waddington et al. 2011, Worrall et al. 2011). However, the porosity of the peat surface on cutover bogs diminishes over time with no intervention due to peat shrinkage and compaction, reducing hydrological conductivity and making moisture unavailable at the surface (Price et al. 2003, Zając et al. 2018). This makes the surface hostile to seed germination and establishment of bog vegetation, particularly non-vascular plants (Renou-Wilson et al. 2019) and Sphagnum mosses (Quinty & Rochefort 2003). Additionally, there are chemical changes (Wind-Mulder et al. 1996) as increased humification and recalcitrance of organic matter reduces nutrient availability to support both plant and microbial communities (Andersen et al. 2006). Selfregeneration to a functioning and sustainable peatland is, therefore, highly unlikely (Money & Wheeler 1999, Quinty & Rochefort 2003) and intervention with active restoration, including plant re-introduction, is needed to re-establish ecohydrological functions and peat accumulation (Rochefort 2000, Chirino et al. 2006, Lucchese et al. 2010).

Natural peatlands in many areas are designated for conservation, and collection of *Sphagnum* (for introduction to restoration sites) is undesirable and prohibited (Gahlert *et al.* 2012, Caporn *et al.* 2018). *Sphagnum* is also slow to establish naturally and effective colonisation would take many years, especially in areas such as bare, extracted peatlands where few local sources exist. Therefore, rapid and sustainable methods of propagation are needed for

effective restoration in necessarily short timescales. Micropropagation Services Ltd (trading as BeadaMoss®) produce large quantities of Sphagnum from tiny amounts of wild-sourced material, using standard tissue-culture techniques involving plant division in a sterile, controlled environment to produce a range of products for different applications (Caporn et al. 2018). BeadaGelTM contains strands of developing Sphagnum suspended in a hydrocolloidal gel, which is applied directly to the peatland surface aiming for even areal coverage. This product contains a mixture of species from five Sphagnum Sections (Acutifolia, Cuspidata, Sphagnum, Squarrosa and Subsecunda), selected to thrive in a range of microhabitats (Atherton et al. 2010, Laine et al. Thus. 2018) across the peatland landscape. Sphagnum has an opportunity to grow wherever the product is placed, as each species is adapted to a particular environmental niche (from pool to hummock) where its productivity is greater than that of other species (Clymo & Hayward 1982).

In establishment phases of bog restoration, vascular plants can 'nurse' and promote Sphagnum growth by providing scaffolding, moss environmental protection and а beneficial microclimate (Ferland & Rochefort 1997. Grosvernier et al. 1997, Quinty & Rochefort 2003, Pouliot et al. 2011) while other mosses such as Polytrichum strictum can reduce plant displacement caused through frost heaving (Price et al. 2003). Eriophorum species are early colonisers, and perhaps species-of-choice to nurse re-colonising the Sphagnum moss (Pouliot et al. 2011, Nugent et al. 2018) as they provide environmental protection and help stabilise the peat surface without out-competing Sphagnum or smothering it with plant litter (Guêné-Nanchen et al. 2017). However, Eriophorum species have aerenchyma, which brings air to the roots and rhizosphere of wetland plants and thus also provides a direct route to the atmosphere for CH₄ from the anaerobic, waterlogged peat (Schimel 1995. Videmšek et al. 2006). This raises concerns that peatlands undergoing restoration may become GHG sources rather than sinks due to elevated CH₄ emissions (Lindsay 2010, Evans et al. 2016) in the short term, until Sphagnum is fully established. On the other hand, oxic conditions created in the rhizosphere of aerenchymatous plants allow methanotrophic bacteria to oxidise CH₄, which distances roots from the methane store (Fritz et al. 2011). On balance, the role of Eriophorum species may be advantageous for climate change mitigation if it facilitates successful peatland restoration.

The aims of this study were to ascertain whether rewetting and planting of *Eriophorum angustifolium*



swards with tissue-cultured *Sphagnum* moss at a degraded lowland bog delivered a carbon greenhouse gas (CGHG) sink or source, whether maturity or type of vegetation were key factors, and to develop a greater understanding of the drivers of gaseous carbon flow to inform future site management. The hypotheses were that:

- 1. restoration of the site will result in a CGHG uptake compared to bare peat;
- 2. CGHG uptake will be greater with maturity of vegetation;
- 3. greater volumes of *E. angustifolium* will result in greater emission of methane;
- 4. the presence of *Sphagnum* moss will reduce the magnitude of methane emission; and
- 5. periods of drought will have a deleterious effect on site CO_2 uptake.

METHODS

Field site

Field trials were conducted on Cadishead Moss (53° 27' 10.8" N, 2° 27' 11.5" W), a UK Site of Biological Importance (SBI) 10 km west-south-west of Manchester, UK (Figure 1) adjacent to peat-extracted Little Woolden Moss. The site is an 8-ha fragment of the once-extensive Chat Moss lowland bog complex which has been mostly urbanised or

drained for agriculture or peat extraction so that only a small fraction of the original complex remains as peatland or under functioning restoration management (Ashby et al. 2021). Cadishead Moss was originally ditch-drained and block-cut for peat, then partially abandoned and colonised by trees and scrub until at least 2000, after which some parts (including the trial areas) were mechanically scraped for peat extraction. The Lancashire Wildlife Trust (LWT) acquired the site in 2009, cleared scrub and trees, and rewetted by installing plastic piling and a series of peat bunds, leaving some deep internal ditches remaining. There is now good coverage of Eriophorum angustifolium in wetter areas and Molinia caerulea in drier areas, with minimal bare peat.

BeadaGel[™] mix

The commercial BeadaGelTM mix provided by the BeadaMoss® company for our field trials contained eleven *Sphagnum* species in the following proportions: *S. auriculatum* Schimp. (synonym: *S. denticulatum*) ~1%, *S. capillifolium* (Ehrh.) Hedw. (ssp. *capillifolium*) ~10%, *S. cuspidatum* Ehrh. Ex Hoffm. ~10%, *S. fallax* (H. Klinggr.) H. Klinggr. ~25%, *S. fimbriatum* Wilson ~10%, *S. medium* Limpr. (likely) (originally designated as *S. magellanicum* Brid.) ~1%, *S. papillosum* Lindb ~10%, *S. tenellum* (Brid.) Bory ~1%, *S. subnitens* Russow & Warnst. ~5%.



Figure 1. Location map of Cadishead and Little Woolden Mosses, close to the conurbation of Manchester, UK. Cadishead Moss and surrounding landscape matrix: map from EDINA Digimap Ordnance Survey Service https://digimap.edina.ac.uk/aerial.



S. magellanicum is now recognised as a species specific to the Southern Hemisphere, and European species similar to *S. magellanicum* are separated into *S. medium* and *S. divinum* (Hassel *et al.* 2018) which have some morphological differences and are generally found, respectively, in ombrotrophic and minerotrophic habitats (Laine *et al.* 2018). The *Sphagnum* sourced for the BeadaMoss® material could be either or a mixture of these, but *S. medium* is assumed. A few strands of each species were

<image>

Figure 2. Examples of collars in the five treatments: a) mature *E. angustifolium* with established BeadaGelTM *Sphagnum* (MEAS); b) as a) without *Sphagnum* (MEA); c) immature, open sward of *E.angustifolium* with new BeadaGelTM *Sphagnum* application (IEAS); d) as c) without *Sphagnum* (IEA); and e) bare peat (BARE). sourced from the Peak District National Park, apart from *S. medium* and *S. tenellum* which were sourced from Cumbria (Caporn *et al.* 2018).

Field plots

Permanent collars were installed in areas of mature and immature *Eriophorum angustifolium*, with and without *Sphagnum*, and bare peat (i.e., five treatments) (Figure 2). All collars were contained within an area of approximately 0.4 ha. Collars were





cut from 300 mm internal diameter plastic waste pipe to 100 mm length and eased into the peat, leaving 40– 50 mm standing above the surface. The commercial BeadaGelTM mix (see previous sub-section) was applied at a rate of 3 L m⁻² to 1 m² areas with immature, open swards of *Eriophorum angustifolium* (with collars in situ) to allow the gel to reach the peat surface. Preliminary CGHG measurements were made after several months, and regular measurements after one year, so that decomposition of any vegetation and roots damaged by collar placement did not influence the results (Rowson *et al.* 2013).

Six trial ('immature') plots were established in October 2015 in areas of sparse E. angustifolium growth. Each plot contained one collar of *E. angustifolium* (IEA), one collar in an area of E. angustifolium with new Sphagnum BeadaGelTM application (IEAS), and one collar on bare peat (BARE). There were no areas with Sphagnum only to provide further controls. At this site, earlier trials of planting Sphagnum on bare peat, even with straw mulch, were either not successful or plots were soon colonised by E. angustifolium; Sphagnum is often only successful in a restoration setting when introduced along with a nurse crop (Quinty & Rochefort 2003, Pouliot et al. 2011). BeadaGel™ was slow to establish during the early stages of the trial on 'immature' plots and was reapplied at the same rate six months after the start of GHG measurements, within the collar area only, at which stage removable environmental protection in the form of greenhouse shading mesh was added to simulate straw mulch.

Three further plots (termed 'mature' plots) were set up at the same time on areas established in June 2014 for a previous study, to allow monitoring at a more advanced stage of BeadaGelTM growth in a tall, dense *E. angustifolium* sward. Each 'mature' plot contained two collars within a 1 m² area of *E. angustifolium* plus *Sphagnum* (MEAS) and one collar on *E. angustifolium* without *Sphagnum* (MEA) nearby. There was no bare peat in areas of mature *E. angustifolium*, so bare peat collars are only associated with 'immature' plots. Throughout the trial, any emerging seedlings of vascular plants other than *E. angustifolium* were removed from the experimental plots, and all seedlings were removed from bare plots, to maintain the experimental design.

A plot dipwell was inserted close to each cluster of collars, within 1.5 m of each collar, to monitor water table depth (WTD), and measurements of WTD relative to the peat surface were recorded manually every week when possible, and fortnightly at minimum. Dipwell design followed Allott *et al.* (2009).

Peat cores and peat quality measurements

Peat cores and quality of surface peat within the trial area were examined for insights about the legacy of degradation which may still influence the carbon balance of the site, and what may be done to remedy it. Nine peat cores to 1 m depth were taken with a Russian corer and visually inspected (only).

Before the start of GHG flux field monitoring, three peat samples were collected to 5 cm depth from around each of the trial plots and homogenised to make up one replicate composite sample per plot (n = 9). Fresh well-mixed peat samples (5 g of each) were added to 25 ml of deionised (DI) water, stirred regularly, then electrical conductivity was measured (Jenway 4510 analyser) in the order of sample preparation. Samples were re-stirred and pH measured (Jenway 3510 analyser), leaving the probe in the solution for 30 s. Further fresh peat samples were prepared for extraction of ammonium and nitrate using 1% KCl (as recommended by Allen 1989) for ion chromatography (IC) (Thermo Scientific Dionex AS analyser) and extraction of elements (Ca, Fe, K, Mg and P) using 0.1M EDTA (as recommended by Lo & Yang 1999) for inductively coupled plasma - optical emission spectrometry (ICP-OES) (Thermo Scientific iCAP 6000 Series ICP Spectrometer). Extractable values were seen as more useful determinants of element bioavailability in the peat than total values (Rosenburgh 2015). Peat samples (mean mass 2.5316 \pm 0.0235 g) were put into 100 ml conical flasks, including one blank sample for each extraction, with 25 ml of the appropriate extraction solution, and the flasks were agitated on an orbital shaker for 30 minutes. Samples were filtered through Sartorius[™] Minisart[™] Plus Syringe Filters (0.2 µm) into tubes for analysis (1 ml for IC and a minimum of 10 ml for ICP-OES), discarding the first 5 ml of filtrate to remove any filter contaminants.

The remainder of the fresh samples were weighed, then oven dried overnight at 105 °C and weighed again to find the mass difference to give sample moisture content. Samples were removed for dry analysis and the remainder re-weighed and placed in a muffle furnace at 550 °C for three hours to find the mass difference (loss-on-ignition) from which organic matter and mineral fractions were estimated. Total C and N content were analysed (LECO FP628 elemental analyser) using 0.1513 \pm 0.0006 g (mean sample mass) of dry, ground peat placed into tared aluminium foil cups twisted into capsules, with five calibration capsules prepared in the same way using EDTA LECO calibration 502-092 (mean mass 0.1508 \pm 0.0005 g).



Gas flux and environmental monitoring

CGHG measurements were obtained using a Los Gatos ResearchTM (LGR) 'Ultraportable Greenhouse Gas Analyser (CH₄,CO₂,H₂O)' (manufactured by Los Gatos Research, San Jose, California, USA), which has < 2 ppb precision for CH₄ and < 300 ppb precision for CO₂ at a 1 second measurement rate. A closed chamber system was created using a clear (Perspex) chamber whereby changes in gas uptake or emission due to plant and soil photosynthesis and respiration could be measured in real time. A Perspex extension was used for taller vegetation, with a partially inflated rubber tyre attached to create a good seal between the collar and the chamber and extension, similar to methods used on automated chambers at Mer Bleue bog (Lai et al. 2012). Gases were directed around the chamber via a ring of tubing, pierced at 1 cm intervals and blocked with silicone gel half-way along to prevent gas cycling in the tube. Each end of the tubing ring was attached via more tubing through the chamber to the inlet and outlet ports of the LGR analyser. A small (9V) fan inside the chamber ensured good gas circulation (tested prior to use). The gas temperature was continuously recorded by the LGR analyser.

CGHG fluxes from each collar were recorded fortnightly during the growing season and monthly during winter for two full years from September 2016 to August 2018, i.e. there were 33 monitoring visits. All collars were monitored, in random order, during each visit. Photosynthetically active radiation (PAR; μ mol m⁻² s⁻¹), peat temperature at 5 cm depth and WTD were recorded during measurements, as recommended by Alm *et al.* (2007). A soil temperature probe (Delta-T Devices Ltd) and PAR meter (Skye Instruments Ltd) were attached to a GP1 Delta-T logger, recording measurements at 10 s intervals.

Dark and light measurements were each taken over a 2-minute period (Davidson *et al.* 2016) with the chamber firstly obscured with a blackout cloth, then aerated before a measurement with the chamber uncovered, to obtain measurements of net ecosystem respiration (NER), net ecosystem exchange (NEE) and CH_4 flux.

Volume of vegetation and headspace within collars

The volumes of the Perspex chamber and extension were known constants. The volume of headspace within the collar was variable owing to changes in the volume of vegetation and swelling/shrinkage of the peat. The depth from top of collar to peat or *Sphagnum* surface was measured at six internal peripheral positions after each gas measurement, and the headspace volume within the collar calculated

and added to the chamber/extension volume for flux calculations. If the *Sphagnum* surface was above the top of the collar, its distance from the collar top was measured after each gas measurement at four internal peripheral positions (midway between centre and edge of collar) and at the highest point (usually central) to obtain a mean height above the collar, giving a negative collar headspace volume.

The volume of *E. angustifolium* within collars was calculated from monthly measurements of the number and length of plants (longest leaf; as outlined by Davidson et al. 2016). If there was dense growth (i.e., more than about 40 plants), the plants were counted and 10 % of representative plants at random peripheral and central positions were measured to obtain a mean. Seasonally, E. angustifolium plants from the site (not within collars) were cut, measured (longest leaf), and the volume of each plant determined by water displacement. Separate scatter graphs of length against volume were plotted for nonflowering and flowering/seeding plants, and used in calculating the volume of plants in each collar (spring, autumn and winter $R^2 = 0.7-0.8$; summer $R^2 = 0.54$ (non-flowering) to 0.6 (flowering)). The volume of *E. angustifolium* (assessed monthly) was subtracted from the headspace when calculating CGHG fluxes.

Measurements of *E. angustifolium* and *Sphagnum* volumes were also used to assess above-ground plant competition and any influence of the changing volume of vegetation on CGHG fluxes over time. Thickness of the *Sphagnum* layer within a collar was estimated as the mean of measurements obtained at nine positions by probing to the recolonised peat surface (which was firm) with a narrow, blunt-ended rod. *Sphagnum* volume was then calculated as that of a cylinder with this height and basal area derived from the estimated percentage *Sphagnum* cover within the collar. Care was taken to minimise damage to the *Sphagnum* when taking measurements.

CGHG measured flux data management

 CO_2 and CH_4 measurements were downloaded from the LGR into Excel spreadsheets and fitted to an Excel linear regression model to obtain gradient (slope), R^2 and *p*-values for each 2-minute measurement. Graphs were used to visualise and remove erroneous start or end measurements, as recommended by Evans *et al.* (2016). The maximum number of observations retained per measurement was 124 and the minimum number was 60. Erroneous periods of measurement could be due to test error or, particularly in summer light levels, low CO_2 availability in the chamber due to high uptake by plants, causing a reduction in slope. Regression line



thresholds of $R^2 > 0.7$ and p < 0.05 were applied to screen for acceptable measurements, similar to protocols used by Evans *et al.* (2016). If neither of these conditions was met the measurement was discarded, but as CO₂ and CH₄ gases were measured concurrently, if one measurement met the criteria a system integrity failure was deemed unlikely and both gas measurements were retained. Only 1 % of all flux measurements were discarded.

Fluxes (g m⁻² h⁻¹) were calculated separately for CO_2 and CH_4 using an equation adapted from Dossa *et al.* (2015):

Flux =
$$(\Delta CO_2/t)^* (PV/RT)^* (1/A_s)^* ((44*60*60)/1000)$$
[1]

where *P* (atm) is atmospheric pressure, *V* (m³) is chamber volume, *R* (L atm mol⁻¹ K) is the universal gas constant, *T* (K) is gas temperature, A_s (m²) is surface area within the collar, and 44 (g mol⁻¹) is the molecular weight of CO₂ (substituted with 16 g mol⁻¹ for CH₄ calculations).

Measurements from the dark and light chambers gave Net Ecosystem Respiration (NER) and Net Ecosystem Exchange (NEE), respectively. Gross Primary Productivity (GPP) was calculated as:

$$GPP = NEE - NER$$
[2]

and values used for further analysis of flux data.

The micrometeorological sign convention was adopted, whereby negative fluxes indicate removal from the atmosphere and positive fluxes indicate addition to the atmosphere.

Methane fluxes were calculated from measurements in the dark. CO_2 equivalents of CH_4 were calculated as $GWP_{100} \times 28$ (Myhre *et al.* 2013) as adopted in the 1997 Kyoto Protocol, when calculating CGHG budgets as CO_2e in g m⁻² yr⁻¹.

Statistical analysis of measured data

Measured vegetation and flux data from all treatment groups (MEAS, MEA, IEAS, IEA, BARE; see Figure 2) were tested for normality using Shapiro Wilk tests, and were found to be not normally distributed. A non-parametric test for repeated measurements (Friedman's test) was used to determine any statistically significant difference between groups and post hoc analysis of flux data with Wilcoxon signed-rank tests was conducted with a Bonferroni correction applied, resulting in a significance level set at p < 0.0083 throughout. Data were analysed statistically using IBM SPSS Statistics for Windows (Version 25.0. Armonk, NY: IBM Corp.) and through the data analysis tools in Microsoft Excel (2019).

CGHG flux data modelling

Hourly air temperature (°C), rainfall (mm) and total solar radiation (W m⁻²) datasets for the full period of measurement (01 Sep 2016 to 31 Aug 2018) were provided by the Whitworth Meteorological Observatory (CAS 2020) at the University of Manchester, 14.8 km east-north-east of the study site. Photosynthetically Active Radiation (PAR) (µmol m⁻² s⁻¹) was derived from solar radiation (W m⁻²) assuming PAR (400–700 nm) is 45 % of total solar radiation, using the conversion factor 1 W m⁻² \approx 2.1 µmol m⁻² s⁻¹ (Biggs 1984).

A linear regression between peat temperature (PT) and air temperature (AT) measurements from micrometeorological equipment on the neighbouring Little Woolden Moss (LWM; Figure 1) was applied to the Whitworth Observatory AT data to provide integrated PT data for the period of CGHG measurements. The record for WTD, measured weekly to fortnightly throughout the study period, was infilled assuming linear changes between measurements, to provide an estimated hourly dataset (Alm *et al.* 2007, Renou-Wilson *et al.* 2019).

The measured CGHG flux data were plotted against the measured environmental variables PT, WTD and PAR and the linear regression R^2 values were ranked to determine which variables best explained the measured values of NER, GPP and methane fluxes (i.e., primary, secondary and tertiary drivers). GPP values were made positive for ease of data manipulation, then non-linear regression (usually exponential) equations were fitted between each measured flux and primary driver to create each primary model. For y_1 , exponential regression ($f:x_1$):

$$y_1 = abx_1 \tag{3}$$

where a = coefficient (or y-intercept), b = exponentand $x_1 = \text{one of the independent environmental}$ variables, i.e. PT (°C) or WTD (cm) or PAR (µmol m⁻² s⁻¹). A linear regression may have interpolated the data better, but an exponential equation ensured that no abnormal negative flux values were introduced. Data from this model were subtracted from measured flux data, to leave residual data. Linear regression equations from residual data were fitted with the secondary driver (and subsequently with the tertiary driver, where used); e.g. for y_2 linear regression (residual: x_2)

$$y_2 = mx_2 + c \tag{4}$$

(where m = line gradient and c = co-efficient, or yintercept). These equations were added iteratively to the primary model, checking for goodness of fit with measured flux data at each stage, to create a final model:



$$f_m = abx_1 + [(f - abx_1)x_2 + c]$$
[5]

where f and f_m = measured and modelled fluxes in CO₂e (g m⁻² h⁻¹).

Equations 3 and 4 were applied to hourly environmental data to provide an integrated model for gaseous fluxes for each treatment. Modelled values for GPP were adjusted to zero when PAR was zero. NEE was calculated from NER and GPP (Equation 2). All data were manipulated using data analysis tools in Microsoft Excel (2019).

The amalgamated treatment data were also modelled using a range of other equations from literature, to check the validity of our model. NER was modelled using a Lloyd & Taylor (1994) model, which is based upon an Arrhenius equation, adapted to incorporate both PT and WTD. NER was also modelled using an extension of that model, developed by Rowson *et al.* (2013), which is a peat temperature model but incorporates a range of environmental variables, as described by Creevy et al. (2020). GPP was modelled using equations which all rely on the relationship between GPP and PAR: (α * GPmax * PAR)/(α * PAR + GPmax) (Michaelis & Menten 1913); $(\alpha * PAR)/(1 - (PAR/2000) + (\alpha *$ PAR/GPmax)) (Falge et al. 2001); and: (a * PAR * GPmax)/sqrt(GPmax² + (α * PAR)²)) (Smith (1936). The alpha constants were derived using Microsoft Excel Solver.

An annual CGHG flux value for each treatment was calculated from the sum of hourly values for each modelled flux dataset for each full year of study (September to August). CO₂ equivalents of CH₄ were calculated by multiplying methane values by a Global Warming Potential (GWP)₁₀₀ of 28 and adding to NEE (balance of NER and GPP), to give a CGHG budget in CO₂e (g m⁻² yr⁻¹). Datasets were analysed for normality using PAST software and found to be not normally distributed through Shapiro-Wilk tests (cross-referenced with Jarque-Bera tests due to large datasets: n = 8760). Differences in fluxes between years within treatment groups were analysed using Microsoft Data Analysis two-sample tests assuming unequal variances. Differences in CGHG flux between treatment groups for each year were tested using PAST software (Kruskal-Wallis with post-hoc Mann-Whitney pairwise test).

RESULTS

Water table depth and meteorological data

Despite higher-than-average rainfall in the preceding three seasons, low rainfall during May to August of Year 2 resulted in a prolonged drought accompanied by a dramatic summer drop in water table to -48 ± 5.9 and -54 ± 6.9 cm (lowest mean value, relative to ground surface) in mature and immature plots respectively, which was 2.1 and 1.7 times (respectively) the largest WTDs recorded during the summer of Year 1 (Figure 3). Long-term average (LTA) data for the area, sourced from the UK Met Office, showed that rainfall was more variable in Year 2 than in Year 1, and in both years was generally higher than average in cooler months and lower than average in warmer months.

Vegetation data

In both years, the volume of *E. angustifolium* in MEAS and IEAS collars (Mature and Immature *E. angustifolium* with *Sphagnum*, respectively) increased rapidly from April and reduced from October, with greater proportional increase in growth in IEAS than MEAS plots. *Sphagnum* volume in both MEAS and IEAS plots levelled off from October/November each year. During the growing season, *Sphagnum* volume increased in MEAS collars in Year 1 but gradually decreased in Year 2, whereas in IEAS collars, *Sphagnum* volume increased throughout, although at a reduced rate during the Year 2 summer drought.

The volume of *E. angustifolium* in both MEA and IEA collars (Mature and Immature *E. angustifolium*only, respectively) changed similarly over the study period, increasing from April and reducing from October (more quickly in IEA collars), but levelled off during the Year 2 Summer drought (reducing slightly in IEA collars).

There was a statistically significant difference in the volume of *E. angustifolium* between MEAS and MEA plots: $\chi^2(2) = 16.860$, p < 0.001, df = 1 as determined by a Friedman Test, (MEA > MEAS). Moreover, in MEAS plots, E. angustifolium volume reduced slightly in Year 2, whereas in MEA plots it increased by 23.5 %. There was a statistically significant difference in the volume of *E. angustifolium* between IEAS and IEA plots: $\chi^2(2)$ = 20.211, p < 0.001, df = 1 as determined by a Friedman Test, with an increasingly greater volume in IEAS than in IEA plots; by 21.2 % in Year 1 and by 42.3 % in Year 2.

Sphagnum volume (zero values in IEAS Year 1 removed) increased overall from Year 1 to Year 2, in MEAS plots by 37.8 % and in IEAS plots by 314 %. However, during the Year 2 summer drought there was obvious drying of vegetation. *Sphagnum* became bleached and the action of taking measurements on collars containing mature *Sphagnum* (which had grown above the top of the collar) isolated plants within the collar from the surrounding *Sphagnum*



carpet. This created a dry edge effect which was less pronounced in collars of immature vegetation, where *Sphagnum* was shorter than the collar and benefitted from mesh shading. Peat shrank in BARE treatments and in some collars of immature *E. angustifolium*, creating a gap between peat and collar.

Peat cores and peat surface samples analysis

Peat cores were visibly different in terms of peat colour, texture and plant species content but had dark and coarse or open-textured (oxidised) peat at the surface, to a depth of 10–28 cm, containing coarse stems and roots of *E. angustifolium*. In most cores there was friable peat at the surface, to a depth of 3–24 cm. Peat colour then changed to variously striated mixtures of black/orange with varying openness of texture, and fine *E. angustifolium* roots throughout. The surface peat characteristics appear typical for a lowland bog (Table 1) although NO₃ and NH₄ levels were high.

Measured flux data

The closed chamber system with the Los Gatos analyser delivered accurate results (94 % above the 0.7 R^2 threshold criteria), with very few (1 %) discarded measurements (Table 2). Reporting of flux

measurements is by replicates of chamber measurements for each treatment across the study period or for each study year (September to August). Measured fluxes of CO_2 and CH_4 (Figure 4) followed the seasonal pattern of rising towards higher summer temperatures and plant and microbial activity, and falling towards lower winter temperatures, plant senescence and reduced microbial activity.

All fluxes from mature vegetation were greater (p < 0.001 throughout) and more variable than those from immature vegetation (mature vegetation: NER 0.018 to 1.031 and GPP -0.019 to -1.91 g m⁻² h⁻¹; CH₄ 0.3 to 2.7 mg m⁻² h⁻¹; immature vegetation: NER 0.014 to 0.51 and GPP -0.020 to -0.87 g m⁻² h⁻¹; CH₄ 0.4 to 1.2 mg m⁻² h⁻¹), and the large range in flux values from each collar can clearly be seen in Figure 4. In mature vegetation, uptake of CO_2 , respiration and CH₄ emissions were greater in plots without Sphagnum, which had greater abundance of vascular plants. In immature vegetation, respiration and CH₄ emissions were similar, but uptake of CO₂ was greater in plots with Sphagnum (which had lower abundance of vascular plants). NER and CH₄ emissions were greater in vegetated plots than bare plots, but there was an overall CO₂ uptake on vegetated plots. Bare plots had some GPP (mean (SE):



Figure 3. Monthly rainfall, long term average (LTA) rainfall, and corresponding mean water table depth (WTD) below the surface, measured across treatment plots grouped by vegetation maturity; study period September 2016 to end-August 2018. LTA (1981 to 2010) rainfall (monthly mean) from Woodford Meteorological Station (53° 20' 24.0" N, 2° 09' 14.4" W), 20 km south-east of the site.



(SE): -0.05 (0.004) g m⁻² h⁻¹) due to algal growth and patches of acrocarpous mosses, but NER was the strongest flux, resulting in CO_2 emission for much of the study period. Methane fluxes were low

throughout, with mean (SE) values of 1.01 (0.05) mg m⁻² h⁻¹ for Mature vegetation, 0.80 (0.02) mg m⁻² h⁻¹ for Immature vegetation, and 0.12 (0.008) mg m⁻² h⁻¹ for BARE.

Table 1. Site characteristics and surface peat properties. Climate data (Met Office 2023) are ten-year (1991–2020) average values from Woodford Observing Site (53° 20' 24.0" N, 2° 09' 14.4" W; 88 m a.s.l.), 23 km SE of Cadishead Moss. Study period 01 Sep 2016 to 31 Aug 2018. Peat properties: mean values (n = 9). Total C and N analysed on LECO Element analyser; EDTA extractable P, K, Ca, Mg analysed on ICP-OES after acid digest with 0.1M EDTA; nitrate and ammonium analysed on KCl-extract using Dionex ion chromatography. Concentrations are on a dry weight basis.

Site character	istics	Peat properties					
Site area	8 ha	NH4 ⁺ (mg kg ⁻¹)	3.55±3.41	pН	4.86±0.21		
Elevation	23 m a.s.l.	NO ₃ ⁻ (mg kg ⁻¹)	1.72±4.34	EC (µS m ²)	45.46±4.54		
Average rainfall 868 mm yr ⁻¹		P (mg g ⁻¹)	0.001 ± 0.001	%Moisture	86±3		
Mean annual air temp 9.8 °C		K (mg g ⁻¹)	0.016 ± 0.018	%OM	96±2		
Mean Jan air temp	4.1 °C	Ca (mg g ⁻¹)	3.31±0.81	%N	1.41 ± 0.08		
Mean Jul air temp	16.2 °C	Mg (mg g ⁻¹)	0.55±0.38	%C	50.81±0.95		
Peat depth	2.3±0.3 m	Bulk density (g cm ⁻³)	0.097 ± 0.023	C/N	36.12±2.36		

Table 2. Results of statistical tests between treatment groups for measured flux values. Friedman's test (X^2 , df, sample no.) and post-hoc (Wilcoxon signed-rank tests Z) tests with statistical significance p < 0.0083. NER = Net Ecosystem Respiration, GPP = Gross Primary Productivity, NEE = Net Ecosystem Exchange. Treatments: M = Mature, I = Immature; EA = *E. angustifolium*-only; EAS = *E. angustifolium* with *Sphagnum*. *NS* = no statistical significance.

Measured flux		NER	GPP	NEE	methane
All treatments	$X^{2}(4, n = 99)$	287.141	257.130	167.064	223.055
	р	< 0.001	< 0.001	< 0.001	< 0.001
	Ζ	-6.949	-6.070	-4.904	-2.841
MEA : MEAS	р	< 0.001	< 0.001	< 0.001	0.004
IEA : IEAS	Ζ	NC	-5.993	-5.993	NC
	р	183	< 0.001	< 0.001	183
	Ζ	-11.633	-12.178	-11.785	-12.091
MEAS DAKE	р	< 0.001	< 0.001	< 0.001	< 0.001
MEA : BARE	Ζ	-8.638	-8.628	-8.195	-8.638
	р	< 0.001	< 0.001	< 0.001	< 0.001
	Ζ	-11.002	-12.050	-11.889	-12.194
IEAS : DAKE	р	< 0.001	< 0.001	< 0.001	< 0.001
IEA : BARE	Ζ	-11.329	-11.785	-11.243	-12.063
	р	< 0.001	< 0.001	< 0.001	< 0.001





Figure 4. Measured Net Ecosystem Respiration (NER) (above) and Gross Primary Productivity (GPP) (below) flux values for each measurement visit (dates are shown on the horizontal axis) in (a) mature and (b) immature plots. MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*); MEA = mature vegetation (*E. angustifolium* only); IEAS = immature vegetation (*E. angustifolium* with *Sphagnum*); BARE = bare peat. In box plots, crosses indicate the mean value, lines indicate the median, and interquartile range is exclusive.





Figure 4 (continued). Measured Net Ecosystem Exchange (NEE) (above) and methane (below) flux values for each measurement visit (dates are shown on the horizontal axis) in (a) mature and (b) immature plots. MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*); MEA = mature vegetation (*E. angustifolium* only); IEAS = immature vegetation (*E. angustifolium* with *Sphagnum*); BARE = bare peat. In box plots, crosses indicate the mean value, lines indicate the median, and interquartile range is exclusive.



Measured flux data and environmental variables

Relationships between the primary driver peat temperature at 5 cm depth (PT) and the secondary driver water table depth (WTD) appeared to explain most of the variability in NER and methane fluxes, and PT followed by PAR then WTD explained variability in GPP (Table 3). GPP appeared to increase until the peat temperature was 16–17 °C, then reduce above this temperature, particularly in immature vegetation where PT was notably higher (maximum 23.5 °C) than in plots with mature vegetation (maximum 19.3 °C). Methane flux increased with greater volumes of *E. angustifolium* ($R^2 = 0.2553$ for Mature, 0.3838 for Immature) but there was no apparent relationship between volume of *Sphagnum* and magnitude of any flux.

Modelled flux data

Modelling of data by treatment, using our model and a range of other models from the literature, produced higher NER R^2 and lower SSE values for the Rowson *et al.* (2013) model in comparison with our own and Lloyd & Taylor + WTD models (Table 4). In contrast, GPP R^2 and SSE values for literature models (Michaelis-Menten, Falge, Smith) were generally poorer than for our model. Annual NER flux values derived from our model and Rowson *et al.* (2013) were very similar, and lower than the fluxes predicted by Lloyd & Taylor + WTD (Table 5). However, for annual GPP flux results, our model generally produced higher values than literature models (although not for BARE treatment) but graphs of modelled and measured fluxes showed a better and

Table 3. R^2 values, coefficients (Standard Error) of linear regression to test relationships between measured environmental variables and fluxes prior to modelling data. MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*); MEA = mature vegetation (*E. angustifolium* only); IEAS = immature vegetation (*E. angustifolium* only); IEAS = immature vegetation (*E. angustifolium* only); BARE = bare peat; NER = net ecosystem respiration; GPP = gross primary productivity; NEE = net ecosystem exchange; PT = peat temperature (°C); WTD = water table depth (cm); PAR = photosynthetically active radiation (μ mol m⁻² s⁻¹). Reported as collated collar values for each treatment. (Data points = 199 other than MEA = 100.)

			MEAS	MEA	IEAS	IEA	BARE
NER	PT	\mathbb{R}^2	0.6418	0.7210	0.5853	0.5822	0.5331
		Coeff (SE)	0.0406 (0.0022)	0.0609 (0.0038)	0.0261 (0.0016)	0.0241 (0.0015)	0.0130 (0.0009)
		\mathbb{R}^2	0.3428	0.4450	0.4087	0.4644	0.4921
	WTD	Coeff (SE)	0.0097 (0.0009)	0.0157 (0.0018)	0.0082 (0.0007)	0.0082 (0.0006)	0.0050 (0.0004)
		\mathbb{R}^2	0.5206	0.6632	0.4524	0.3102	0.1021
GPP	РТ	Coeff (SE)	-0.0715 (0.0049)	-0.1171 (0.0085)	-0.0453 (0.0036)	-0.0326 (0.0035)	-0.0029 (0.0006)
	PAR	\mathbb{R}^2	0.2923	0.3179	0.2989	0.2534	0.1081
		Coeff (SE)	-0.0004 (4.97E-05)	-0.0007 (9.96E-05)	-0.0003 (3.82E-05)	-0.0003 (3.50E-05)	-3.08E-05 (6.33E-06)
	WTD	\mathbb{R}^2	0.0889	0.2176	0.1961	0.1378	0.0645
		Coeff (SE)	-0.0097 (0.0022)	-0.0220 (0.0042)	-0.0112 (0.0016)	-0.0083 (0.0015)	-0.0009 (0.0002)
		\mathbb{R}^2	0.0694	0.1130	0.1323	0.0406	0.3181
METHANE (as CO ₂ e)	РТ	Coeff (SE)	0.0013 (0.0003)	0.0024 (0.0007)	0.0007 (0.0001)	0.0005 (0.0002)	0.0003 (3.17E-05)
		\mathbb{R}^2	0.0000	0.0026	0.0116	0.0113	0.3295
	WTD	Coeff (SE)	-3.97E-06 (1.11E-04)	0.0001 (0.0002)	7.61E-05 (5.03E-05)	9.80E-05 (6.58E-05)	0.0001 (1.25E-05)



consistent fit for our GPP model compared to those from literature (Figures A1–A4 in the Appendix).

Modelled and measured values (Figure 5) showed similar trends although modelled GPP had high extrapolated points during peak-temperature periods, particularly in Mature vegetation treatments. In Year 1, Modelled winter flux values appeared to be greater than measured values, although the values were small, but generally the remainder of the measured fluxes were within the range of modelled values. There was high variability in measured values of methane fluxes, which the model smoothed to follow the measured mean, although modelled winter values were a little higher than measured values in Year 1.

Table 4. Comparison of R^2 (SSE: sum of squared estimate of errors) values obtained for linear regressions between measured and variously modelled data for NER and GPP by treatment groups. MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*), MEA = mature vegetation (*E. angustifolium* only), IEAS = immature vegetation (*E. angustifolium* with *Sphagnum*), IEA = immature vegetation (*E. angustifolium* only) and BARE = bare peat control plots.

R ² values (SSE)		NER		GPP				
	This Study	Lloyd & Taylor (1994) + WTD	Rowson <i>et al.</i> (2013)	This Study	Michaelis & Menten (1913)	Falge <i>et al</i> . (2001)	Smith (1936)	
MEAS	0.55 (4.55)	0.65 (4.03)	0.72 (2.58)	0.47 (19.33)	0.30 (24.68)	0.30 (24.67)	0.30 (24.71)	
MEA	0.61 (4.46)	0.73 (3.40)	0.82 (1.64)	0.48 (24.74)	0.33 (25.14)	0.33 (38.79)	0.32 (25.33)	
IEAS	0.46 (4.08)	0.59 (2.69)	0.65 (2.22)	0.29 (19.77)	0.33 (16.79)	0.32 (17.00)	0.32 (50.30)	
IEA	0.45 (3.68)	0.59 (2.43)	0.67 (1.97)	0.23 (14.91)	0.28 (13.37)	0.28 (13.37)	0.28 (13.32)	
BARE	0.38 (1.61)	0.57 (0.83)	0.61 (0.75)	0.18 (0.38)	0.12 (2.20)	0.12 (2.26)	0.12 (5.05)	

Table 5. Comparison of annual CO₂ flux values (g m⁻² yr⁻¹) obtained through variously modelled data for NER and GPP by treatment groups (all collar data within each group). MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*), MEA = mature vegetation (*E. angustifolium* only), IEAS = immature vegetation (*E. angustifolium* only) and BARE = bare peat control plots.

Annual modelled flux comparisons			NER		GPP				
		This Study	Lloyd & Taylor (1994) + WTD	Rowson <i>et al.</i> (2013)	This Study	Michaelis & Menten (1913)	Falge <i>et al.</i> (2001)	Smith (1936)	
MEAS	Year 1	2082.34	2457.44	2067.44	-2622.53	-1359.65	-1347.09	-1315.20	
	Year 2	2067.68	2449.82	2065.86	-2231.26	-1431.94	-1420.70	-1389.69	
MEA	Year 1	3190.40	3448.65	3169.61	-4107.46	-2068.49	-1226.75	-1982.67	
	Year 2	3191.75	3461.93	3165.34	-3644.63	-2178.29	-1288.91	-2097.31	
IEAS	Year 1	1055.66	1431.60	1207.14	-1570.75	-1195.43	-1226.75	-1982.67	
	Year 2	1071.74	1524.63	1237.25	-1495.38	-1244.09	-1288.91	-2097.31	
	Year 1	1097.26	1403.56	1192.52	-1237.02	-956.60	-956.60	-922.03	
IEA	Year 2	1121.77	1479.55	1216.15	-1192.39	-998.45	-998.45	-964.86	
BARE	Year 1	451.74	639.18	531.08	-198.69	-364.49	-367.50	-478.70	
	Year 2	460.17	710.10	587.54	-203.94	-382.36	-385.70	-505.68	





Figure 5. Modelled and measured Net Ecosystem Respiration (NER) (Rowson *et al.* 2013 model) and Gross Primary Productivity (GPP) (This Study model) on treatments a) MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*), b) MEA = mature vegetation (*E. angustifolium* only), Lines show modelled hourly data, points show mean measurements \pm SE.





Figure 5 (continued). Modelled and measured Net Ecosystem Respiration (NER) (Rowson *et al.* 2013 model) and Gross Primary Productivity (GPP) (This Study model) on treatments c) IEAS = immature vegetation (*E. angustifolium* with *Sphagnum*), d) IEA = immature vegetation (*E. angustifolium* only), Lines show modelled hourly data, points show mean measurements \pm SE.

Figure 5 (continued). Modelled and measured Net Ecosystem Respiration (NER) (Rowson *et al.* 2013 model) and Gross Primary Productivity (GPP) (This Study model) on treatment e) BARE plots = bare peat control plots. Modelled and measured CH₄-CO₂e on f) mature plots. Lines show modelled hourly data, points show mean measurements \pm SE.

Figure 5 (continued). Modelled and measured CH₄-CO₂e on g) immature plots. Lines show modelled hourly data, points show mean measurements \pm SE.

Modelled CGHG budgets

Between study years, annual NER in vegetated plots (Table 6) was similar for each treatment (IEAS p <0.05; other treatments no significant difference (NS)). Both GPP (IEAS p < 0.01, IEA p < 0.05, other treatments p < 0.001) and consequently NEE (p <0.001 throughout) were lower in Year 2, and methane flux converted to CO₂e also (p < 0.001 throughout). When CH₄-CO₂e flux was added to NEE to give an overall annual carbon greenhouse gas (CGHG) budget, there was a reduction in the Year 2 CGHG uptake in MEAS, MEA and IEAS plots (p < 0.001), and an increase in emission in IEA (p < 0.001) plots. In BARE plots, NER was higher in Year 2 (p < 0.001) and GPP was similar in both years (p NS) which increased NEE in Year 2 (p < 0.001); CH₄-CO₂e was also higher in Year 2 (p < 0.001) resulting in increased CGHG emission in Year 2 compared to Year 1 (p < 0.001). Methane emission from BARE plots was 11-12 % of that from MEAS, IEAS and IEA plots, and 6 % of that from MEA plots.

The difference in CGHG budgets between treatment groups (Table 6) was statistically significant in both Year 1 (H (4) = 301.1, p < 0.001) and Year 2 (H (4) = 661.2, p < 0.001). Post-hoc

Mann-Whitney pairwise tests showed statistically significant differences between all treatments (p < 0.001) apart from MEA and both IEA and BARE (p NS) in Year 1.

CGHG uptake was larger in plots of mature vegetation without *Sphagnum* (MEA>MEAS) in both years (p < 0.001) and larger in plots of immature vegetation with *Sphagnum* (IEAS>IEA) (both years p < 0.001). In all plots without *Sphagnum*, uptake was higher in mature than in immature vegetation throughout (MEA>IEA) (p NS in Year 1; p < 0.001 in Year 2). In all plots with *Sphagnum*, uptake was higher in mature vegetation in Year 1 (MEAS> IEAS) (p < 0.001) but higher in immature vegetation in year 2 (IEAS>MEAS) (p < 0.001).

The data showed an overall progression towards a CGHG sink function in this peatland restoration site during Year 1 (Figure A5), with high CGHG emission from bare peat and a net CGHG emission becoming a net uptake as vegetation colonised and matured. However, the drier conditions during Year 2 reduced CGHG uptake significantly, particularly in mature vegetation, and resulted in CGHG emission from plots of mature vegetation with *Sphagnum*.

A.T. Keightley et al. ENGINEERING RESTORATION AND GASEOUS C UPTAKE ON A DEGRADED BOG

Table 6. Treatment composition and characteristics with associated modelled annual sum flux measurements. MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*); MEA = mature vegetation (*E. angustifolium* only); IEAS = immature vegetation (*E. angustifolium* with *Sphagnum*); IEA = immature vegetation (*E. angustifolium* only); WTD = water table depth (cm); NER = net ecosystem respiration; GPP = gross primary productivity; NEE = net ecosystem exchange; CO₂ equivalents of CH₄ were calculated by multiplying by Global Warming Potential (GWP)₁₀₀ of 28 and added to NEE values to give a CO₂e CGHG budget in g m⁻² yr⁻¹ (final column); values other than fluxes reported as means of measurements \pm SD.

Treatment	Year	WTD (cm)	Eriophorum angustifolium (cm ³)	Sphagnum (cm ³)	NER (g m ⁻² yr ⁻¹)	GPP (g m ⁻² yr ⁻¹)	NEE (g m ⁻² yr ⁻¹)	Methane (g m ⁻² yr ⁻¹)	Methane as CO ₂ e (g m ⁻² yr ⁻¹)	$\begin{array}{c} CO_2e\\ CGHG \ budget\\ (g \ m^{-2} \ yr^{-1}) \end{array}$
	1	5.88 ± 6.45	212.63 ± 126.29	5968.39 ± 2567.12	2067.44	-2622.53	-555.09	7.16	200.46	-354.63
MEAS	2	11.77 ± 17.21	214.27 ± 106.25	8223.62 ± 2851.15	2065.86	-2231.26	-165.40	6.62	185.45	20.05
	(mean)	9.30 ± 14.05	213.54 ± 115.02	7221.30 ± 2940.18	2066.65	-2426.89	-360.24	6.89	192.95	-167.29
	1	5.88 ± 6.45	302.84 ± 132.25	0	3169.61	-4107.46	-937.85	13.66	382.53	-555.32
MEA	2	11.77 ± 17.21	371.73 ± 114.66	0	3165.34	-3644.63	-479.30	12.78	357.77	-121.52
	(mean)	9.30 ± 14.05	341.11 ± 126.41	0	3167.47	-3876.05	-708.57	13.22	370.15	-338.42
1	1	18.70 ± 14.47	68.39 ± 49.22	310.59 ± 157.31	1207.14	-1570.75	-363.61	6.66	186.52	-177.09
IEAS	2	18.69 ± 15.36	117.19 ± 70.20	975.72 ± 429.50	1237.25	-1495.38	-258.13	6.49	181.67	-76.47
	(mean)	18.69 ± 14.92	95.50 ± 66.14	822.23 ± 475.78	1222.19	-1533.07	-310.87	6.57	184.09	-126.78
	1	18.70 ± 14.47	55.63 ± 42.58	0	1192.52	-1237.02	-44.50	7.11	199.08	154.59
IEA	2	18.69 ± 15.36	82.37 ± 50.78	0	1216.15	-1192.39	23.76	7.03	196.86	220.62
	(mean)	18.69 ± 14.92	70.48 ± 48.95	0	1204.34	-1214.70	-10.37	7.07	197.97	187.61
BARE	1	18.70 ± 14.47	0	0	531.08	-198.69	332.39	0.75	21.02	353.42
	2	18.69 ± 15.36	0	0	587.54	-203.94	383.60	0.78	21.73	405.33
	(mean)	18.69 ± 14.92	0	0	559.31	-201.31	358.00	0.76	21.38	379.37

DISCUSSION

In Year 1 this study showed a progression from continual CGHG emission from bare peat, through a reduction in emissions on initial colonisation with bog vegetation (in this case Eriophorum angustifolium), to a large CGHG uptake by mature recolonising E. angustifolium, and a benefit from introduction of Sphagnum mosses to immature vegetation. However, in Year 2 with variable weather patterns and a long period of summer drought, the CGHG uptake was small, and areas of bare peat and sparse vegetation, particularly with no Sphagnum cover, showed increasing emissions of CGHG, and there was a change from CGHG uptake to emission in the mature E. angustifolium with Sphagnum treatment. This could suggest that the site has minimal resilience to anticipated climate change in the UK such as the increased frequency of hot summers (Lowe et al. 2018, Met Office 2019). However, although drought in the second year may have limited the potentially positive influence of increasing vegetation cover, the avoided CGHG losses through restoring the site (i.e., not leaving it bare) were large, so supporting hypothesis 1, that restoration of the site will result in a CGHG uptake compared to bare peat. This highlights the urgent nature of restoring degraded peatlands for best outcomes in terms of climate change mitigation targets (Nugent et al. 2019).

One of the aims of this study was to ascertain the influence of the type and maturity of vegetation on CGHG fluxes, but altered weather patterns between the two years of study influenced volume and condition of both E. angustifolium and Sphagnum, and thus their capacity for CGHG uptake. E. angustifolium growth and senescence followed expected seasonal trends in the first year of study, but growth stalled in the second summer in all plots but IEAS, indicating that the drought may have caused early senescence in the vascular plants (Bubier *et al.* 2003), and reduced their photosynthesis. The Year 2 summer drought also reduced Sphagnum volume in mature plots and caused severe surface desiccation, reducing photosynthetic potential (McNeil & Waddington 2003, Bortoluzzi et al. 2006, Rydin & Jeglum 2013, Helfter et al. 2014). Additionally, lower PAR in the spring of Year 2 (probably cloudier conditions) may have also limited photosynthesis (Lafleur et al. 2003, Loisel et al. 2012) and contributed to lower GPP for the whole year (Nijp et al. 2015).

Conversely, *Sphagnum* in immature plots grew in volume throughout the study period, with mesh shading to support early establishment and some

environmental protection provided through being lower in the E. angustifolium sward than Sphagnum within mature E. angustifolium. This may have helped retain soil moisture and reduce evapotranspiration for a healthy layer of Sphagnum, favour but also appeared to continued E. angustifolium growth and development in these plots during the drought period. Mesh shading was used to replicate straw mulching but is not likely to be employed in large-scale restoration works, where straw-mulching is routinely used.

In mature vegetation, the volume of E. angustifolium was less in plots with Sphagnum than without, which may be related to reduction in exposure of E. angustifolium leaves to sunlight as the Sphagnum grew and increasingly covered them, or a greater capacity of Sphagnum to harvest nutrients for growth (Malmer et al. 2003, Bragazza et al. 2004, Fritz et al. 2014). Immature Sphagnum had not reached a height at which it competed with E. angustifolium for light. The water table was higher in plots with mature vegetation than those with immature vegetation and bare peat. Wilson et al. (2013) also found this, and potential explanations are that evaporation is reduced through increased shade from dense E. angustifolium (Price et al. 2003), or that a higher water table supports proliferation of *E. angustifolium* (Rochefort et al. 2016). Topographical differences between collar locations were not measured, but may have influenced plant growth.

CO₂ uptake through GPP was greater in plots of mature E. angustifolium only, which had the greatest volume of E. angustifolium overall, suggesting that Hypothesis 2, CGHG uptake will be greater with maturity of vegetation, was supported. But the overall picture was more complex. NER emission was also highest in these plots, and greater than in plots with immature vegetation or bare peat, and rates of NER and GPP were closely related. This accords with studies suggesting NER is higher in vegetated than in BARE plots and is related to litterfall, temperature and rainfall (Bortoluzzi et al. 2006, Evans et al. 2016, Jordan et al. 2016), but the most important factor may be inputs of carbon from photosynthesis (Bond-Lamberty et al. 2004). However, greater NER emission was also related to warmer, drier conditions, which concurs with most literature sources (e.g., Danevčič et al. 2010, Wang et al. 2014), particularly for bare peat (Bortoluzzi et al. 2006) although this can alter depending on the plant assemblage in a heterogeneous peatland system (Juszczak et al. 2013). The NER increase in BARE plots each year could be related to the encroachment of surrounding vegetation, and root growth within the

column of bare peat inside the collar, even though surface vascular plant growth was removed, but is more likely due to greater microbial decomposition in drier, warmer conditions (Juszczak *et al.* 2013). Algae and acrocarpous bryophytes (no *Sphagnum* growth) were not removed, however, as this is a natural progression of bare peat cover, and this may have influenced carbon cycling in these plots.

Methane emissions (as CO₂e) considerably reduced CGHG uptake in each vegetated treatment, highlighting the importance of including methane in GHG studies (Bussell et al. 2010, Haddaway et al. 2014). However, methane fluxes were generally midrange to low in all treatments compared to some studies on rewetted sites using instantaneous measurements (e.g., Beyer & Höper 2015, Davidson et al. 2016, Evans et al. 2016), which is perhaps to be expected in a site where microbial communities are still recovering from the effects of long-term drainage during peat cutting and subsequent evapotranspiration from scrub cover (Andersen et al. 2013, Juottonen et al. 2015, Nugent et al. 2018) and perhaps a reduction in substrate nutrient availability (Basiliko et al. 2007) prior to restoration ten years ago. It might be expected that methane fluxes in this study would increase as the site matures (in the short term) with more plant growth, plant litter and a higher, more stable water table, leading to greater availability of labile carbon (Glatzel et al. 2004, Lafleur et al. 2005, Urbanová et al. 2011): measured seasonal results in this study prior to the Year 2 summer drought supported that trend. MEA plots had volumes of E. angustifolium greater and approximately twice the methane emission of other vegetated plots, supporting Hypothesis 3, that greater volumes of E. angustifolium will result in greater emission of methane. However, overall methane fluxes in the second year with drought were lower throughout vegetated plots compared to the previous wet year, in accordance with the accepted view that methane flux declines (CH₄ oxidises) in dry sites (Danevčič et al. 2010, Turetsky et al. 2014, Abdalla et al. 2016). There was no particular relationship between high water table and high methane emissions in vegetated plots overall, concurring with other studies, e.g., Wilson et al. (2022), who found the main driver of CH₄ flux on a rewetted site to be soil temperature, but contrary to earlier empirical evidence (Glatzel et al. 2004, Danevčič et al. 2010, Urbanová et al. 2011, Evans et al. 2016).

Methane flux was higher in vegetated plots without *Sphagnum*, even in immature vegetation where *E. angustifolium* volume was higher in plots with *Sphagnum* than without, supporting Hypothesis 4, that the presence of *Sphagnum* moss will reduce the magnitude of methane emission. Flux from bare plots was 9 % that of vegetated plots overall. These results concur with Bortoluzzi et al. (2006) who reported the rank of highest to lowest fluxes to be Eriophorum-dominated, Sphagnum-dominated, then bare plots, and Couwenberg et al. (2011) who found a strong relationship between methane flux and the density of aerenchymatous leaves. Methane flux be related to the amount appears to of E. angustifolium, but is also reduced when Sphagnum present, suggesting some methanotrophic is consumption of methane in the Sphagnum layer (Kip et al. 2010, Larmola et al. 2010, van Winden et al. 2012, Nugent et al. 2018).

In common with Leppälä et al. (2011), this study found that reduction in CGHG uptake in a dry year compared to the previous wet year was driven more by changes in GPP than NER. However, plant dynamics were a complex factor. Greater volumes of E. angustifolium appeared to be related to greater CGHG uptake. This is partly contrary to findings of Kivimäki et al. (2008) that stands of mixed sedges and Sphagnum sequestered more carbon than those of sedges alone due to lower NER, but the maturity of the vegetation is a factor in our study. Wilson *et al.* (2013) found greater NEE in plots with sedges when compared to those with *Sphagnum* only. Moreover, Tuittila et al. (1999) suggested that a restored site colonised with mature *Eriophorum* (*E. vaginatum*) was a carbon sink resilient to interannual changes in weather. However, other studies found that NEE was greater in rewetted Sphagnum-dominated than in sedge-dominated sites (Bever & Höper 2015, Evans et al. 2016, Renou-Wilson et al. 2019) due to a lower NER to GPP ratio, and the presence of Sphagnum may reduce NER by retaining moisture (Waddington & Warner 2001). In our study, there was a lower NER to GPP ratio each year in E. angustifolium-only than with Sphagnum in mature stands, but the reverse was so in immature stands, with the lowest NER to GPP ratio overall each year in immature E. angustifolium with Sphagnum. An open sward of immature E. angustifolium emitted CGHG, increasingly so in the drought year, but with a layer of Sphagnum it became a CGHG sink each year.

Lower CGHG emission through NER and higher uptake through GPP in immature plots with *Sphagnum* than without suggests an effect of both increased moisture retention (Waddington & Warner 2001) and more plant material on the peat substrate. This may indicate that establishment of a layer of *Sphagnum* is more crucial in immature than in mature vegetation, in terms of CGHG uptake, and so efforts to create a beneficial microclimate at the peat surface (e.g., mulch, nurse planting, etc.) should be one of the

Mires and Peat, Volume 29 (2023), Article 23, 33 pp., http://www.mires-and-peat.net/, ISSN 1819-754X

fundamental processes for initial stages of peatland restoration (Quinty & Rochefort 2003, Groeneveld *et al.* 2007, Waddington *et al.* 2010, Pouliot *et al.* 2011). Early micropropagated mixed-species *Sphagnum* introduction into *E. angustifolium* also appears to create a mutually supportive environment for both plants, whereas attempting to introduce it when *E. angustifolium* is already well-established does not seem to promote good *Sphagnum* outcomes, particularly in dry conditions.

The large range in collar flux measurement within treatments (noted by Bortoluzzi et al. (2006) in their study, and seen in data from other sources, e.g., Wilson et al. 2013, Beyer & Höper 2015, Renou-Wilson et al. 2019), demonstrates the heterogeneity of the site, the complex nature of associations between carbon cycling in degraded peatlands under restoration measures and fluctuations in environmental factors, and perhaps that using vegetation type and density as a proxy for carbon balance measurements may not capture the CGHG state of individual sites sufficiently.

This study found the E. angustifolium-dominated area of Cadishead Moss with and without Sphagnum introduction to be an overall net CGHG sink, despite the methane emissions, in Year 1 and an overall net CGHG source in Year 2, which included a summer drought. The mean (SE) CO2e CGHG emission of vegetated monitoring points was -233.11 (150.56) g m⁻² yr⁻¹ in Year 1 and 10.67 (75.96) g m⁻² yr⁻¹ in Year 2. This supported Hypothesis 5, that periods of drought, which generate a low water table, will have a deleterious effect on site CO₂ uptake. The mean (SE) yearly CO₂e CGHG emission from bare peat of 379.37 (25.96) g m⁻² yr⁻¹ shows the benefits of restoration in terms of avoided CGHG losses (Worrall et al. 2011, Renou-Wilson et al. 2019). The CO₂e sink strength in vegetated plots in Year 1 is in the mid-range of some other similar restored bogs (e.g., those studied by Drewer et al. 2010, Beyer & Höper 2015 and Renou-Wilson et al. 2019). The findings from this study are contrary to those of Evans et al. (2016) at the nearby Astley Moss (rewetted cut-over bog), where measurements were taken on a generally inundated part of the site, and very high methane emissions (43.7 g m⁻² yr⁻¹) pushed the CO₂e NEE sink of -336 g m⁻² yr⁻¹ (revised to -41 g m⁻² yr⁻¹ across the entire site based on vegetation assemblage) into a site CGHG source. More data are needed on lowland bogs under restoration to further refine the inventory of greenhouse gas emission factors for UK peatlands (Evans et al. 2017), and our study can contribute to that.

This study site is not yet in equilibrium, has a widely fluctuating WTD, and changed over the study

period in terms of vegetation cover and density. Few studies, assessing GHG fluxes in relation to type of vegetation (Strack & Zubak 2013, Wilson *et al.* 2013, Renou-Wilson *et al.* 2019), and over time (Waddington *et al.* 2010), fully address the question of the dynamic nature of vegetation on fluxes in a single peatland system under restoration measures over time, although there are some good, recent examples (e.g., Nugent *et al.* 2018, Nugent *et al.* 2019). This broader, integrated approach is worth exploring in more depth, particularly when funding for restoration work may depend on evidence for change over short timescales.

The study area had high peat NO₃ and NH₄ levels (Table 1), which could be related to intermittent aerobic conditions (Urbanova et al. 2011), but Cadishead Moss is close to the large conurbation of Greater Manchester, so may have legacy pollution effects from the Industrial Revolution (Garcés-Pastor et al. 2023) as well as current effects from regional oxidised and reduced nitrogen pollution. Modelled nitrogen deposition to the site is approximately 18.9 kg ha⁻¹ yr⁻¹, well above empirical critical loads for nitrogen for lowland raised bogs of 5-10 kg ha⁻¹ yr⁻¹ (UKCEH 2023). Low water table is of particular concern, as the influence of nitrogen deposition is increased when concentrated in solution (Pearce & Van der Wal 2008) and there may be cumulative N-load effects (Sheppard et al. 2014), which compromise bryophyte growth, increase algal growth (Payne et al. 2013), and make scrub development more likely on a dry peatland site with high atmospheric nitrogen deposition (Krupa 2003, Bubier et al. 2007).

Significant proportions of the top 50 cm of peat cores taken were black, oxidised, rough-textured peat, showing evidence of the compaction and hydrological instability typical of damaged peatlands where the acrotelm has been removed (Price et al. 2003, Lindsay & Clough 2016). Moreover, all bare plots, and some plots with immature vegetation, cracked during the summer drought, which is a likely sign of humification, and no doubt allowed greater evaporation down the peat profile (Lindsay & Clough 2016). Because this area of the site has been previously mechanically scraped with none of even original block-cut surface remaining, the rehabilitation to ecohydrological function, particularly in the short term, is questionable (Price et al. 2003). Long-term degradation of the site has resulted in poor-quality surface and sub-surface peats, which are likely to contribute to poor hydrological control and continue to create obstacles to restoration and reduced capacity for CGHG uptake. Chemical analysis of the changing

environment on this degraded site over time, particularly with integrated studies of surface peats, plants and water, could be used to quantify restoration progress (Andersen *et al.* 2010), and would allow greater understanding of the nutrient cycling and microbial activity underpinning its capacity for CGHG sequestration. However, particular focus should be concentrated on improving hydrological control to maintain higher WTD during dry periods to promote long-term recovery.

If climate change continues to affect weather patterns, summer drought may become more common (Lowe *et al.* 2018, Met Office 2019). If so, it is likely that the CGHG sink function of peatlands under restoration will reduce unless greater resilience, particularly in terms of maintaining water table levels to support *Sphagnum* proliferation, can be engineered so that a functional acrotelm redevelops in the long term.

ACKNOWLEDGEMENTS

This article was derived from a doctoral thesis (PhD) part-funded by Micropropagation Services (EM) Ltd, match-funded by Manchester Metropolitan University (MMU). The authors thank David McKendry and technical officers at MMU for their support; Dr Roxanne Anderson, University of the Highlands and Islands and Dr Robert Sparkes, Manchester Metropolitan University, who examined the PhD thesis and provided valuable comments and corrections; and Lancashire Wildlife Trust for site access permissions.

AUTHOR CONTRIBUTIONS

ATK co-originated and planned the work, undertook all field and laboratoratory work, wrote the drafts and final paper, and is the lead author; CDF, JGR and SJMC had academic oversight, co-originated the work, assisted in planning and installing field equipment, and commented on all drafts; NAW cooriginated the work, supplied and applied *Sphagnum* materials, assisted in planning, and commented on drafts.

REFERENCES

Abdalla, M., Hastings, A., Truu, J., Espenberg, M., Mander, Ü, Smith, P. (2016) Emissions of methane from northern peatlands: a review of management impacts and implications for future management options. *Ecology and Evolution*, 6, 7080–7102.

- Aitova, E., Morley, T., Wilson, D., Renou-Wilson, F. (2023) A review of greenhouse gas emissions and removals from Irish peatlands. *Mires and Peat*, 29, 04, 17 pp.
- Allen, S.E. (ed.) (1989) Chemical Analysis of Ecological Materials. Second edition, Blackwell Scientific Publications, Oxford, 368 pp.
- Allott, T.E.H., Evans, M.G., Lindsay, J.B., Agnew, C., Freer, J.E., Jones, A., Parnell, M. (2009) Water Tables in Peak District Blanket Peatlands. Report No. 17, Moors for the Future, Edale, 49 pp.
- Alm, J., Shurpali, N.J., Tuittila, E-S., Laurila, T., Maljanen, M., Saarnio, S., Minkkinen, K. (2007) Methods for determining emission factors for the use of peat and peatlands - flux measurements and modelling. *Boreal Environment Research*, 12, 85–100.
- Andersen, R., Francez, A.-J., Rochefort, L. (2006) The physicochemical and microbiological status of a restored bog in Québec: Identification of relevant criteria to monitor success. *Soil Biology* & *Biochemistry*, 38, 1375–1387.
- Andersen, R., Rochefort, L., Poulin, M. (2010) Peat, water and plant tissue chemistry monitoring: a seven-year case-study in a restored peatland. *Wetlands*, 30, 159–170.
- Andersen, R., Chapman, S.J., Artz, R.R.E. (2013) Microbial communities in natural and disturbed peatlands: A review. *Soil Biology & Biochemistry*, 57, 979–994.
- Ashby, M., Zini, V., Holt, A. (2021) *Chat Moss Natural Capital Assessment*. Peel L&P commissioned report, Natural Capital Solutions Ltd., Towcester, 52 pp.
- Atherton, I., Bosanquet, S., Lawley, M. (2010) Mosses and Liverworts of Britain and Ireland: A Field Guide. British Bryological Society, Plymouth, 848 pp.
- Bain, C.G., Bonn, A., Stoneman, R., Chapman, S. and 21 others (2011) *IUCN UK Commission of Inquiry on Peatlands*. IUCN UK Peatland Programme, Edinburgh, 109 pp.
- Basiliko, N., Blodau, C., Roehm, C., Bengtson, P., Moore, T.R. (2007) Regulation of decomposition and methane dynamics across natural, commercially mined, and restored northern peatlands. *Ecosystems*, 10, 1148–1165.
- Beyer, C., Höper, H. (2015) Greenhouse gas exchange of rewetted bog peat extraction sites and a *Sphagnum* cultivation site in northwest Germany. *Biogeosciences*, 12, 2101–2117.
- Biggs, W.W. (1984) *Principles of Radiation Measurement*. LI-COR® Lincoln, NE, 8 pp.

- Bond-Lamberty, B., Wang, C., Gower, S.T. (2004) A global relationship between the heterotrophic and autotropic components of soil respiration? *Global Change Biology*, 10, 1756–1766.
- Bortoluzzi, E., Epron, D., Siegenthaler, A., Gilbert, D., Buttler, A. (2006) Carbon balance of a European mountain bog at contrasting stages of regeneration. *New Phytologist*, 172, 708–718.
- Bragazza, L., Tahvanainen, T., Kutnar, L., Rydin, H., Limpens, J., Hájek, M., Grosvernier, P., Hájek, T., Hájková, P., Hansen, I., Iacumin, P., Gerdol, R. (2004) Nutritional constraints in ombrotrophic *Sphagnum* plants under increasing atmospheric nitrogen deposition in Europe. *New Phytologist*, 163, 609–616.
- Bubier, J.L., Bhatia, G., Moore, T.R., Roulet, N.T., Lafleur, P.M. (2003) Spatial and temporal variability in growing-season net ecosystem carbon dioxide exchange at a large peatland in Ontario, Canada. *Ecosystems*, 6, 353–367.
- Bubier, J.L., Moore, T.R., Bledzki, L.A. (2007) Effects of nutrient addition on vegetation and carbon cycling in an ombrotrophic bog. *Global Change Biology*, 13(6), 1168–1186.
- Bussell, J., Jones, D.L., Healey, J.R., Pullin, A. (2010) How do draining and re-wetting affect carbon stores and greenhouse gas fluxes in peatland soils? (systematic review). Collaboration for Environmental Evidence, Review 08-012 (SR49). Online at: www.environmentalevidence.org/SR49.html, accessed 22 Nov 2023.
- Caporn, S.J.M., Rosenburgh, A.E., Keightley, A.T., Hinde, S.L., Riggs, J.L., Buckler, M., Wright, N.A. (2018) *Sphagnum* restoration on degraded blanket and raised bogs in the UK using micropropagated source material: a review of progress. *Mires and Peat*, 20, 09, 17 pp.
- CAS (2020) *The Whitworth Meteorological Observatory*. Web page, Centre for Atmospheric Science (CAS), The University of Manchester. Online at: http://www.cas.manchester.ac.uk/ restools/whitworth/, accessed 22 Nov 2023.
- Chirino, C., Campeau, S., Rochefort, L. (2006) *Sphagnum* establishment on bare peat: The importance of climatic variability and *Sphagnum* species richness. *Applied Vegetation Science*, 9, 285–294.
- Clymo, R.S., Hayward, P.M. (1982) The ecology of *Sphagnum*. In: Smith, A.J.E. (ed.) *Bryophyte Ecology*, Chapman and Hall, London, 229–289.
- Couwenberg, J., Thiele, A., Tanneberger, F., Augustin, J., Bärisch, S., Dubovik, D., Liashchynskaya, N., Michaelis D., Minke, M., Skuratovich, A., Joosten, H. (2011) Assessing

greenhouse gas emissions from peatlands using vegetation as a proxy. *Hydrobiologia*, 674, 67–89.

- Creevy, A.L., Payne, R.J., Andersen, R., Rowson, J.G. (2020) Annual gaseous carbon budgets of forest-to-bog restoration sites are strongly determined by vegetation composition. *Science of the Total Environment*, 705, 135863, 11 pp.
- Crump, J. (ed.) (2017) Smoke on Water: Countering Global Threats from Peatland Loss and Degradation - A Rapid Response Assessment. United Nations Environment Programme (UNEP) and GRID-Arendal, Nairobi and Arendal, 70 pp.
- Danevčič, T., Mandic-Mulec, I., Stres, B., Stopar, D., Hacin, J. (2010) Emissions of CO₂, CH₄ and N₂O from Southern European peatlands. *Soil Biology* & *Biochemistry*, 42, 1437–1446.
- Davidson, S.J., Sloan, V.L., Phoenix, G.K., Wagner, R., Fisher, J.P., Oechel, W.C., Zona, D. (2016) Vegetation type dominates the spatial variability in CH₄ emissions across multiple arctic tundra landscapes. *Ecosystems*, 19, 1116–1132.
- Dossa, G.G.O., Paudel, E., Wang, H., Cao, K., Schaefer, D., Harrison, R.D. (2015) Correct calculation of CO₂ efflux using a closed-chamber linked to a non-dispersive infrared gas analyser. *Methods in Ecology and Evolution*, 6(12), 1435– 1442.
- Drewer, J., Lohila, A., Aurela, M., Laurila, T., Minkkinec, K., Pentilä, T., Dinsmore, K.J., McKenzie, R.M., Helfter, C., Flechard, C., Sutton, M.A., Skiba, U.M. (2010) Comparison of greenhouse gas fluxes and nitrogen budgets from an ombrotrophic bog in Scotland and a minerotrophic sedge fen in Finland. *European Journal of Soil Science*, 61, 640–650.
- Evans, C., Morrison, R., Burden, A., Williamson, J. and 31 others (2016) *Final Report on Project SP1210: Lowland Peatland Systems in England and Wales - Evaluating Greenhouse Gas Fluxes and Carbon Balances.* Centre for Ecology and Hydrology, Bangor, 170 pp.
- Evans, C., Artz, R., Moxley, J., Smyth, M-A., Taylor,
 E., Archer, N., Burden, A., Williamson, J.,
 Donnelly, D., Thomson, A., Buys, G., Malcolm,
 H., Wilson, D., Renou-Wilson, F., Potts, J. (2017) *Implementation of an Emission Inventory for UK Peatlands*. Report to the Department for Business,
 Energy and Industrial Strategy, Centre for
 Ecology and Hydrology, Bangor, 88 pp.
- Evans, C.D., Peacock, M., Baird, A.J., Artz, R.R.E. and 27 others (2021) Overriding water table control on managed peatland greenhouse gas emissions. *Nature*, 593, 548–552.
- Falge, E., Baldocchi, D., Olson, R., Anthoni, P., Aubinet, M., Bernhofer, C., Burba, G., Ceulemans,

R., Clement, R., Dolman, H., Granier, A. (2001) Gap filling strategies for defensible annual sums of net ecosystem exchange. *Agricultural and Forest Meteorology*, 107(1), 43–69.

- Ferland, C., Rochefort, L. (1997) Restoration techniques for *Sphagnum*-dominated peatlands. *Canadian Journal of Botany*, 75(7), 1110–1118.
- Fritz, C., Pancotto, V.A., Elzenga, J.T.M., Visser, E.J.W., Grootjans, A.P., Pol, A., Iturraspe, R., Roelofs, J.G.M., Smolders, A.J.P. (2011) Zero methane emission bogs: extreme rhizosphere oxygenation by cushion plants in Patagonia. *New Phytologist*, 190, 398–408.
- Fritz, C., Lamers, L.P.M., Riaz, M., van den Berg, L.J.L., Elzenga, T.J.T.M. (2014) Sphagnum mosses - masters of efficient N-uptake while avoiding intoxication. *PLoS ONE*, 9(1), e79991, 11 pp.
- Gahlert, F., Prager, A., Schulz, J., Krebs, M., Gaudig, G., Joosten, H. (2012) *Sphagnum* propagules from spores: first experiences. Extended Abstract No. 307, *Proceedings of the 14th International Peat Congress*, International Peat Society, Stockholm, 5 pp.
- Garcés-Pastor, S., Fletcher, W.J., Ryan, P.A. (2023) Ecological impacts of the industrial revolution in a lowland raised peat bog near Manchester, NW England. *Ecology and Evolution*, 13(2), e9807, 22 pp.
- Glatzel, S., Basiliko, N., Moore, T. (2004) Carbon dioxide and methane production potential of peats from natural, harvested and restored sites, Eastern Quebec, Canada. *Wetlands*, 24(2), 261–267.
- Glenk, K., Faccioli, M., Martin-Ortega, J., Schulz, C., Potts, J. (2021) The opportunity cost of delaying climate action: Peatland restoration and resilience to climate change. *Global Environmental Change*, 70, 102323, 14 pp.
- González, E., Rochefort, L. (2014) Drivers of success in 53 cutover bogs restored by a moss layer transfer technique. *Ecological Engineering*, 68, 279–290.
- Gorham, E. (1991) Northern peatlands: role in the carbon cycle and probable responses to climatic warming. *Ecological Applications*, 1(2), 182–195.
- Graver, B., Zhang, K., Rutherford, D. (2019) CO₂ Emissions from Commercial Aviation, 2018.
 Working paper 2019-16, The International Council on Clean Transportation (ICCT), 13 pp. Online at: https://theicct.org/publication/co2emissions-from-commercial-aviation-2018/, accessed 25 Nov 2023.
- Groeneveld, E.V.G., Massé, A., Rochefort, L. (2007) *Polytrichum strictum* as a nurse-plant in peatland

restoration. Restoration Ecology, 15(4), 709–719.

- Grosvernier, P., Matthey, Y., Buttler, A. (1997) Growth potential of three *Sphagnum* species in relation to water table level and peat properties with implications for their restoration in cut-over bogs. *Journal of Applied Ecology*, 34(2), 471– 483.
- Guêné-Nanchen, M., Pouliot, R., Hugron, S., Rochefort, L. (2017) Effect of repeated mowing to reduce graminoid plant cover on the moss carpet at a *Sphagnum* farm in North America. *Mires and Peat*, 20, 06, 12 pp.
- Günther, A., Barthelmes, A., Huth, V., Joosten, H., Jurasinski, G., Koebsch, F., Couwenberg, J. (2020) Prompt rewetting of drained peatlands reduces climate warming despite methane emissions. *Nature Communications*, 11, 1644, 5 pp.
- Haddaway, N.R., Burden, B., Evans, C.D., Healey, J.R., Jones, D.L., Dalrymple, S.E., Pullin, A.S. (2014) Evaluating effects of land management on greenhouse gas fluxes and carbon balances in boreo-temperate lowland peatland systems. *Environmental Evidence*, 3, 5, 30 pp.
- Hassel, K., Kyrkjeeide, M.O., Yousefi, Y., Prestø, T., Stenøien, H.K., Shaw, A.J., Flatberg, K.I. (2018) *Sphagnum divinum* (sp. nov.) and *S. medium* Limpr. their relationship to *S. magellanicum* Brid. *Journal of Bryology*, 40, 197–222.
- Helfter, C., Campbell, C., Dinsmore, K.J., Drewer, J., Coyle, M., Anderson, M., Skiba, U., Nemitz, E., Billett, M.F., Sutton, M.A. (2014) Drivers of longterm variability in CO₂ net ecosystem exchange in a temperate peatland. *Biogeosciences Discussions*, 11, 14981–15018.
- Joosten, H., Tapio-Biström, M.-L., Tol, S. (eds.) (2012) *Peatlands - Guidance for Climate Change Mitigation Through Conservation, Rehabilitation and Sustainable Use.* Second edition, Mitigation of Climate Change in Agriculture 5, FAO and Wetlands International, Rome, 100 pp.
- Joosten, H., Sirin, A., Couwenberg, J., Laine, J., Smith, P. (2016) The role of peatlands in climate regulation. In: Bonn, A., Allott, T., Evans, M., Joosten, H., Stoneman, R. (eds.) *Peatland Restoration and Ecosystem Services: Science, Policy and Practice*, Cambridge University Press, Cambridge, 493 pp.
- Jordan, S., Strömgren, M., Fiedler, J., Lundin, L., Lode, E., Nilsson, T. (2016) Ecosystem respiration, methane and nitrous oxide fluxes from ecotopes in a rewetted extracted peatland in Sweden. *Mires and Peat*, 17, 07, 23 pp.
- Juottonen, H., Kotiaho, M., Robinson, D., Merilä, P., Fritze, H., Tuittila, E. (2015) Microform-related

community patterns of methane-cycling microbes in boreal *Sphagnum* bogs are site specific. *FEMS Microbiology Ecology*, 91(09), fiv094, 13 pp.

- Juszczak, R., Humphreys, E., Acosta. M., Galczewska, M.M., Kayzer, D., Olejnik, J. (2013) Ecosystem respiration in a heterogeneous temperate peatland and its sensitivity to peat temperature and water table depth. *Plant and Soil*, 366(1–2), 505–520.
- Kip, N., van Winden, J.F., Pan, Y., Bodrossy, L., Reichart, G.J., Smolders, A.J.P., Jetten, M.S.M., Sinninghe Damsté, J.S., Op den Camp, H.J.M. (2010) Global prevalence of methane oxidation by symbiotic bacteria in peat-moss ecosystems. *Nature Geoscience*, 3(9), 617–621.
- Kivimäki, S.K., Yli-petäys, M., Tuittila, E.-S. (2008) Carbon sink function of sedge and *Sphagnum* patches in a restored cut-away peatland: increased functional diversity leads to higher production. *Journal of Applied Ecology*, 45, 921–929.
- Krupa, S.V. (2003) Effects of atmospheric ammonia (NH₃) on terrestrial vegetation: a review. *Environmental Pollution*, 124, 179–221.
- Lafleur, P.M., Roulet, N.T., Bubier, J.L., Frolking, S., Moore, T.R. (2003) Interannual variability in the peatland-atmosphere carbon dioxide exchange at an ombrotrophic bog. *Global Biogeochemical Cycles*, 17(2), 1036, 14 pp.
- Lafleur, P.M., Moore, T.R., Roulet, N.T., Frolking, S. (2005) Ecosystem respiration in a cool temperate bog depends on peat temperature but not water table. *Ecosystems*, 8, 619–629.
- Lai, D.Y.F., Roulet, N.T., Humphreys, E.R., Moore, T.R., Dalva, M. (2012) The effect of atmospheric turbulence and chamber deployment period on autochamber CO₂ and CH₄ flux measurements in an ombrotrophic peatland. *Biogeosciences*, 9, 3305–3322.
- Laine, J., Flatberg, K.I., Harju, P., Timonen, T., Minkkinen, K., Laine, A., Tuittila, E.-S., Vasander, H. (2018) Sphagnum Mosses - The Stars of European Mires. Department of Forest Sciences, University of Helsinki, Helsinki, 326 pp.
- Larmola, T., Tuittila, E., Tiirole, M., Nykänen, H., Martikainen, P.J., Yrjälä, K., Tuomivirta, T., Fritze, H. (2010) The role of *Sphagnum* mosses in the methane cycling of a boreal mire. *Ecology*, 91(8), 2356–2365.
- Leifeld, J., Menichetti, L. (2018) The underappreciated potential of peatlands in global climate change mitigation strategies. *Nature Communications*, 9, 1071, 7 pp.
- Leppälä, M., Laine, A.M., Seväkivi, M.-L., Tuittila, E.-S. (2011) Differences in CO₂ dynamics

between successional mire plant communities during wet and dry summers. *Journal of Vegetation Science*, 22, 357–366.

- Lindsay, R. (2010) Peatbogs and Carbon: A Critical Synthesis to Inform Policy Development in Oceanic Peat Bog Conservation and Restoration in the Context of Climate Change. RSPB Scotland, 315 pp.
- Lindsay, R.A., Clough, J. (2016) A Review of the Influence of Ombrotrophic Peat Depth on the Successful Restoration of Bog Habitat. Commissioned Report No. 925, Scottish Natural Heritage, 74 pp.
- Lloyd, J., Taylor, J.A. (1994) On the temperature dependence of soil respiration. *Functional Ecology*, 8(3), 315–323.
- Lo, I.M.C., Yang, X.Y. (1999) EDTA extraction of heavy metals from different soil fractions and synthetic soils. *Water, Air, & Soil Pollution*, 109, 219–236.
- Loisel, J., Gallego-Sala, A.V., Yu, Z. (2012) Globalscale pattern of peatland *Sphagnum* growth driven by photosynthetically active radiation and growing season length. *Biogeosciences*, 9, 2737– 2746.
- Lowe, J.A., Bernie, D., Bett, P., Bricheno, L., Brown, S., Calvert, D., Clark, R., Eagle, K., Edwards, T., Fosser, G., Fung, F. (2018) UKCP18 Science Overview Report. Met Office Hadley Centre, Exeter, 73 pp.
- Lucchese, M., Waddington, J., Poulin, M., Pouliot, R., Rochefort, L., Strack, M. (2010) Organic matter accumulation in a restored peatland: evaluating restoration success. *Ecological Engineering*, 36, 482–488.
- Malmer, N., Albinsson, C., Svensson, B.M., Wallén, B. (2003) Interferences between *Sphagnum* and vascular plants: effects on plant community structure and peat formation. *Oikos*, 100, 469– 482.
- Mazziotta, A., Granath, G., Rydin, H., Bengtsson, F., Norberg, J. (2019) Scaling functional traits to ecosystem processes: Towards a mechanistic understanding in peat mosses. *Journal of Ecology*, 107, 843–859.
- McNeil, P., Waddington, J.M. (2003) Moisture controls on *Sphagnum* growth and CO₂ exchange on a cutover bog. *Journal of Applied Ecology*, 40, 354–367.
- Met Office (2019) *UK Climate Projections: Headline Findings, September 2019, Version 2.* Met Office Hadley Centre, Exeter, 10 pp.
- Met Office (2023) UK climate averages: Woodford (Greater Manchester). Online at: https://www. metoffice.gov.uk/research/climate/maps-and-

data/uk-climate-averages/gcqrqyr80, accessed 23 Nov 2023.

- Michaelis, L., Menten, M.L. (1913) Die Kinetik der Invertinwirkung (The kinetics of invertase action). *Biochemische Zeitschrift*, 49, 334–336 (in German).
- Money, R.P., Wheeler, B.D. (1999) Some critical questions concerning the restoration of damaged raised bogs. *Applied Vegetation Science*, 2, 107–116.
- Myhre, G., Shindell, D., Bréon, F.-M., Collins, W., Fuglestvedt, J., Huang, J., Koch, D., Lamarque, J.-F., Lee, D., Mendoza, B., Nakajima, T., Robock, A., Stephens, G., Takemura, T., Zhang, H. (2013) Anthropogenic and natural radiative forcing. In: *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change* [Stocker, T.F., D. Qin, G.-K. Plattner, M. Tignor, S.K. Allen, J. Boschung, A. Nauels, Y. Xia, V. Bex and P.M. Midgley (eds.)]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 1535 pp.
- Nijp, J.J., Limpens, J., Metselaar, K., Peichl, M., Nilsson, M.B., van der Zee, S.E.A.T.M., Berendse, F. (2015) Rain events decrease boreal peatland net CO₂ uptake through reduced light availability. *Global Change Biology*, 21, 2309– 2320.
- Nugent, K., Strachan, I.B., Strack, M., Roulet, N., Rochefort, L. (2018) Multi-year net ecosystem carbon balance of a restored peatland reveals a return to carbon sink. *Global Change Biology*, 24, 5751–5768.
- Nugent, K.A., Strachan, I.B., Roulet, N.T., Strack, M., Frolking, S., Helbig, M. (2019) Prompt active restoration of peatlands substantially reduces climate impact. *Environmental Research Letters*, 14(12), 124030, 9 pp.
- Payne, R.J., Jassey, V.E.J., Leith, I.D., Sheppard, L.J., Dise, N.B., Gilbert, D. (2013) Ammonia exposure promotes algal biomass in an ombrotrophic peatland. *Soil Biology & Biochemistry*, 57, 936–938.
- Pearce, I.S.K., Van der Wal, R. (2008) Interpreting nitrogen pollution thresholds for sensitive habitats: the importance of concentration versus dose. *Environmental Pollution*, 52, 253–256.
- Pouliot, R., Rochefort, L., Karofeld, E., Mercier, C. (2011) Initiation of *Sphagnum* moss hummocks in bogs and the presence of vascular plants: Is there a link? *Acta Oecologica*, 37, 346–354.
- Price, J.S., Heathwaite, A.L., Baird, A.J. (2003) Hydrological processes in abandoned and restored

peatlands: An overview of management approaches. *Wetlands Ecology and Management*, 11, 65–83.

- Quinty, F., Rochefort, L. (2003) *Peatland Restoration Guide: Second Edition.* Canadian *Sphagnum* Peat Moss Association, St. Albert AB and New Brunswick Department of Natural Resources and Energy, Fredericton NB, 106 pp.
- Renou-Wilson, F., Moser, G., Fallon, D., Farrell, C.A., Müller, C., Wilson, D. (2019) Rewetting degraded peatlands for climate and biodiversity benefits: Results from two raised bogs. *Ecological Engineering*, 127, 547–560.
- Rochefort, L. (2000) *Sphagnum* A keystone genus in habitat restoration. *The Bryologist*, 103(3), 503–508.
- Rochefort, L., LeBlanc, M.C., Bérubé, V., Hugron, S., Boudreau, S., Pouliot, R. (2016) Reintroduction of fen plant communities on a degraded minerotrophic peatland. *Botany*, 94(11), 1041–1051.
- Rosenburgh, A.E. (2015) *Restoration and Recovery* of Sphagnum on Degraded Blanket Bog. Doctoral dissertation, Manchester Metropolitan University, Manchester, 238 pp.
- Rowson, J.G., Worrall, F., Evans, M.G. (2013) Predicting soil respiration from peatlands. *Science of the Total Environment*, 442, 397–404.
- Rydin, H., Jeglum, J.K. (2013) *The Biology of Peatlands*. Second edition, Oxford University Press, Oxford, 382 pp.
- Schimel, J.P. (1995) Plant transport and methane production as controls on methane flux from arctic wet meadow tundra. *Biogeochemistry*, 28, 183–200.
- Sheppard, L.J., Leith, I.D., Mizunuma, T., Leeson, S., Kivimaki, S., Neil Cape, J., van Dijk, N., Leaver, D., Sutton, M.A., Fowler, D., Van den Berg, L.J., Crossley, A., Field, C., Smart, S. (2014) Inertia in an ombrotrophic bog ecosystem in response to 9 years' realistic perturbation by wet deposition of nitrogen, separated by form. *Global Change Biology*, 20(2), 566–580.
- Smith E.L. (1936) Photosynthesis in relation to light and carbon dioxide. *Proceedings of the National Academy of Sciences (PNAS)*, 22(8), 504–511.
- Strack, M., Zuback, Y.C.A. (2013) Annual carbon balance of a peatland 10 yr following restoration. *Biogeosciences*, 10, 2885–2896.
- Tuittila, E.-S., Komulainen, V.-M., Vasander, H., Laine, J. (1999) Restored cut-away peatland as a sink for atmospheric CO₂. *Oecologia*, 120, 563– 574.
- Turetsky, M.R., Kotowska, A., Bubier, J., Dise, N.B., Crill, P., Hornibrook, E.R.C., Minkkinen, K.,

Moore, T.R., Myers-Smith, I.H., Nykänen, H., Olefeldt, D., Rinne, J., Saarnio, S., Shurpali, N., Tuittila, E., Waddington, M., White, J.R., Wickland, K.P., Wilmking, M. (2014) A synthesis of methane emissions from 71 northern, temperate, and subtropical wetlands. *Global Change Biology*, 20, 2183–2197.

- UKCEH (2023) Air Pollution Information System (APIS). UK Centre for Ecology & Hydrology (UKCEH). Online at: https://www.apis.ac.uk/ search-location, accessed 28 Jun 2023.
- Urbanová, Z., Picek, T., Bárta, J. (2011) Effect of peat re-wetting on carbon and nutrient fluxes, greenhouse gas production and diversity of methanogenic archaeal community. *Ecological Engineering*, 37, 1017–1026.
- van Breemen, N. (1995) How Sphagnum bogs down other plants. Trends in Ecological Evolution (TREE), 10, 270–275.
- van Winden, J.F., Reichart, G.J., McNamara, N.P., Benthien, A., Damsté, J.S.S. (2012) Temperatureinduced increase in methane release from peat bogs: a mesocosm experiment. *PLoS ONE*, 7(6), e39614, 5 pp.
- Videmšek, U., Turk, B., Vodnik, D. (2006) Root aerenchyma - formation and function. *Acta Agriculturae Slovenica*, 87(2), 445–453.
- Waddington, J.M., Warner, K.D. (2001) Atmospheric CO₂ sequestration in restored mined peatlands. *Écoscience*, 8(3), 359–368.
- Waddington, J.M., Strack, M., Greenwood, M.J. (2010) Toward restoring the net carbon sink function of degraded peatlands: Short-term response in CO₂ exchange to ecosystem-scale restoration. *Journal of Geophysical Research*, 115, G01008, 13 pp.
- Waddington, J.M., Lucchese, M.C., Duval, T.P. (2011) *Sphagnum* moss moisture retention

following the re-vegetation of degraded peatlands. *Ecohydrology*, 4, 359–366.

- Wang, X., Liu, L., Piao, S., Janssens, I.A., Tang, J., Liu, W., Chi, Y., Wang, J., Xu, S. (2014) Soil respiration under climate warming: differential response of heterotrophic and autotrophic respiration. *Global Change Biology*, 20, 3229– 3237.
- Wilson, D., Farrell, C., Mueller, C., Hepp, S., Renou-Wilson, F. (2013) Rewetted industrial cutaway peatlands in western Ireland: a prime location for climate change mitigation? *Mires and Peat*, 11, 01, 22 pp.
- Wilson, D., Mackin, F., Tuovinen, J-P., Moser, G., Farell, C., Renou-Wilson, F. (2022) Carbon and climate implications of rewetting a raised bog in Ireland. *Global Change Biology*, 28, 6349–6365.
- Wind-Mulder, H.L., Rochefort, L., Vitt, D.H. (1996) Water and peat chemistry comparisons of natural and post-harvested peatlands across Canada and their relevance to peatland restoration. *Ecological Engineering*, 7, 161–181.
- Worrall, F., Chapman, P., Holden, J., Evans, C., Artz, R., Smith, P., Grayson, R. (2011) A Review of Current Evidence on Carbon Fluxes and Greenhouse Gas Emissions from UK Peatland. JNCC Report No. 442, JNCC, Peterborough, 86 pp.
- Yu, Z.C. (2012) Northern peatland carbon stocks and dynamics: a review. *Biogeosciences*, 9(10), 4071.
- Zając, E., Zarzycki, J., Ryczek, M. (2018) Degradation of peat surface on an abandoned post-extracted bog and implications for revegetation. *Applied Ecology and Environmental Research*, 16(3), 3363–3380.

Submitted 01 Apr 2023, revision 26 Sep 2023 Editor: Olivia Bragg

Author for correspondence:

Dr Chris Field, Senior Lecturer in Environmental Ecology, Faculty of Science & Engineering, Department of Natural Sciences, John Dalton Building, Chester Street, Manchester, M1 5GD, UK. Tel: +44 (0)161 247 6203; Email: c.field@mmu.ac.uk

Figure A1. Graphs showing goodness of fit between mean measured and variously modelled data for NER by treatment groups. Values measured (at intervals of two weeks to one month) are shown by points with error bars (SE), superposed on a graph of hourly values modelled using the methods of This Study, Rowson *et al.* (2013) or Lloyd & Taylor (1994) + WTD. The treatments are: MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*) and MEA = mature vegetation (*E. angustifolium* only).

Figure A2. Graphs showing goodness of fit between mean measured and variously modelled data for NER by treatment groups. Values measured (at intervals of two weeks to one month) are shown by points with error bars (SE), superposed on a graph of hourly values modelled using the methods of This Study, Rowson *et al.* (2013) or Lloyd & Taylor (1994) + WTD. The treatments are: IEAS = immature vegetation (*E. angustifolium* with *Sphagnum*), IEA = immature vegetation (*E. angustifolium* only) and BARE = bare peat control plots.

Figure A3. Graphs showing goodness of fit between mean measured and variously modelled data for GPP by treatment groups. Values measured (at intervals of two weeks to one month) are shown by points with error bars (SE), superposed on a graph of hourly values modelled using the methods of This Study, Michaelis & Menten (1913), Falge *et al.* (2001) or Smith (1936). The treatments are: MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*) and MEA = mature vegetation (*E. angustifolium* only).

Figure A4. Graphs showing goodness of fit between mean measured and variously modelled data for GPP by treatment groups. Values measured (at intervals of two weeks to one month) are shown by points with error bars (SE), superposed on a graph of hourly values modelled using the methods of This Study, Michaelis & Menten (1913), Falge *et al.* (2001) or Smith (1936). The treatments are: IEAS = immature vegetation (*E. angustifolium* with *Sphagnum*), IEA = immature vegetation (*E. angustifolium* only) and BARE = bare peat control plots.

Figure A5. Modelled yearly fluxes comparing treatments. Top row: NER, GPP and NEE in Year 1 and Year 2; bottom left: methane as CO_2e in Years 1 and 2; and bottom right: CGHG balance as CO_2e in Years 1 and 2. The treatments are: MEAS = mature vegetation (*E. angustifolium* with *Sphagnum*); MEA = mature vegetation (*E. angustifolium* only); IEAS = immature vegetation (*E. angustifolium* with *Sphagnum*); Bare = bare peat. Shared letters below each flux line indicate no significant statistical difference between treatments.

